ORIGINAL ARTICLE



Waste seeds of *Mangifera indica*, *Artocarpus heterophyllus*, and *Schizizium commune* as biochar for heavy metal removal from simulated wastewater

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Abstract

The threat of arsenic contamination in water is a challenging issue worldwide. Millions of people utilize untreated groundwater having high levels of arsenic in developing countries. Design Expert 6.0.8 has been used to design experiments and carried out statistical analysis for optimization of different parameters. It is of prime importance to develop cheap environment friendly bio-sorbent for protecting health of the poor from ill effects of arsenic. In the present investigation, we prepared bio-sorbent from the solid waste seed biomass of *Mangifera indica* (M), *Artocarpus heterophyllus* (JF), and *Schizizium commune* (JP). The characterization of bio-sorbents has been done by using different techniques namely FTIR and XRD. Arsenic concentration was estimated using ICP and adsorption parameters optimized for pH, adsorbent dose, and initial arsenic concentration. At pH 8.4, kinetics study of arsenic removal was M (94%), JF (93%), and JP (92%) for initial concentration of 2.5 ppm. The adsorption kinetics was well explained by Freundlich model and pseudo-second reaction order.

Keywords Arsenic removal · Bio-adsorbent · Schiziziumcommun · Mangiferaindica · Biochar

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1 Introduction

Presence of arsenic in the form of trivalent arsenite (As III) and pentavalent arsenate (As V) in the terrestrial and aquatic habitats is threatening human health seriously. As per World Health organization, arsenic is group one human carcinogen [1]. In surface water, the forms of arsenic are transformed from their inorganic to partially organic behavior due to biological activity. Arsenic is naturally present in the water of across the globe including the USA, Mexico, Argentina, India, and Bangladesh [2]. In addition to natural presence of it, some anthropogenic activities also enhance its percentage in soil and water. Apart from natural source of arsenic, many industries like mining, cement production, and fossil fuel burning are producing it. The use of arsenic based herbicides and pesticides pollute agriculture field [3, 4]. The arsenic present in agriculture field on the plant surface may reach to the human body directly with improperly washed food surface. Alternatively, it may enter in aquatic water bodies with runoff or percolate to groundwater and enters in the human body with drinking water and cause ill effects to human in long term [5]. However, most of the environmental arsenic problems are due to natural sources. Predominant forms of arsenic in the toxic condition of running water and pond is pentavalent arsenic acid and oxyanionic forms, whereas, in the anoxic or moderately reducing conditions of groundwater it exists in more stable trivalent arsenite (As III). Predominant form of arsenic is determined the pH value. In the natural water bodies of pH ranging from 5 to 8, the pentavalent form is about five times compared to trivalent form [6]. The groundwater arsenic concentration may be high up to the level of 3500 ppm, and its toxicity affected population is more than 200 million globally [7]. The concentration of arsenic has been frequently reported high in different water bodies across the world. Considering the toxic and carcinogenic nature, a 10-ppm concentration of arsenic in drinking is the maximum permissible limit [27].

Efforts have been made to remove arsenic from polluted water bodies employing techniques like coagulation, flocculation, membrane filtration, reverses osmosis, precipitation, bio-remediation, and adsorption. These water treatment technologies are continuously being updated to achieve the aim of easy, efficient, and cost-effective treatment of arsenic polluted water [28]. Nanomaterials have been synthesized from iron oxides and copper nano-flakes activated carbons as low cast alternatives [8]. The nano-materials with unique porous structure and high chemical and mechanical resistance are useful, and they are efficiently employed for the purpose [9-11]. The powdered nano-adsorbent adsorbs metal from wastewater and leaving clean water on the top of the vessel.

Variety of materials has been tested as adsorbent and adsorption behavior in the including stones, shells, and even sewage products, in recent past. However, the use of sewage product could give undesirable effects on the human body due to relatively less knowledge about the microorganisms or their enzymes present [12, 13]. To reduce the adsorbents cost significantly wastes from the food industry and agriculture has been utilized, and these materials are generally described as bioadsorbent. Wastes of agriculture and food industry are potential bio-sorbent for arsenic removal from water. Bio-sorbents have also been prepared from the seeds of the plant and shown to be appropriate raw materials for bio-sorbents [14], [29]. Biochemically, bio-sorbents are composed of carbohydrate, lipid/fat, proteins, lignin, cellulose, and hemicelluloses materials in varying proportion depending upon the source plant. These compounds contain functional groups like hydroxyl, carboxyl, amide, amine, and carbamide that are able to facilitate complex formation and removal of arsenic from the aqueous solution [15].

Considering the need of cheap, efficient, eco-friendly, and potentially harmless bio-adsorbent, in the present investigation, we prepared easily available seed kernels of *Mangifera indica* (Mango), *Artocarpus heterophyllus* (Jack fruit, JF), and *Schizizium commune* (Java Plum, JP) for synthesizing bio-sorbents. In this current COVID 19 pandemic situation, the demand of fresh water increases tremendously. Most of the population in developing countries are depends of groundwater for their need. In the condition of increasing demand more hand pumps and bore wells are required in various regions so the chances of exposure of arsenic contaminated water increases continuously.

The aim of the current investigation was achieved for an efficient arsenic removal technique by using low-cost biomass adsorbent. The home-made adsorbent has been prepared and characterized using various techniques like FTIR and XRD. The experiments were performed for the duration of 120 min and sample was collected at a 10-min interval to check the kinetics. The experiments were performed by changing pH, amount dose, and initial arsenic composition. The arsenic adsorption data of selected bio-adsorbents was well supported by spectroscopic data, adsorption isotherms, and kinetic models.

2 Materials and method

2.1 Experimental procedure

Batch adsorption experiment performed as explained elsewhere [8]. In brief, a experiment is performed in a 50-ml beaker containing arsenic solution and adsorbent continuously stirred by a magnetic stirrer with minimum of 5 rpm. Different aqueous arsenic concentration (0.8, 1, 1.5, 2, and 2.5 ppm) were exposed separately to various doses (20, 40, 60, 80, and 100 mg) of adsorbent at a pH (5 to 11) for optimizing the adsorption experiment. The experiments performed for 2 h and samples were withdrawn at the regular interval of 10 min and filtered with a 0.45μ -filter before analysis for residual arsenic concentration in the solution using ICP.

The equilibrium test was performed for understanding arsenic sorption on bio-sorbent. Dry adsorbent (0.5 g/L) was added in the flasks containing a 50 ml arsenic solution (pH=7). The initial arsenic concentration of solution tested for adsorption range from 5 to 300 mg/L. Arsenic solutions with bio-sorbents were stirred for 24 h and then bio-sorbents were filtered with 0.45 μ filters. The equilibrium arsenic concentration, Ce (mg/L), was determined by analyzing the filtrate by ICP. The sorption capacity (mg. g⁻¹) of bio-sorbent at equilibrium is obtained using Equation given below:

$$q_e = \frac{(C_{0-}C_e)V}{m} \tag{1}$$

2.2 Plants used for bio-sorbent

All the angiospermic plants used in the present study are deciduous perennial trees. They are distributed across the India in different climatic zones. The *Mangifera indica* L. is a very well-known economic fruit plant belonging to family Anacardiaceae; their fruits are irregularly egg-shaped. *Syzygium cumini* commonly called Jamun belongs to the family Myrtaceae. The plant has been well documented for its antidiabetic and hypoglycemic effects in its seed, fruit, and bark. Artocarpus heterophyllus is an evergreen tree of about 10–20 m that belongs to family Moraceae. The fruit contains 100–300 seeds $(10-12 \times 8-10 \text{ mm})$, about 15–18% of the fruit weight. The seeds are good source of starch and protein. The seed starch has high tolerance for thermal and mechanical shear.

2.3 Preparation of bio-sorbent

Seeds of different fruits like mango, jackfruit, and jamun have been collected from campus of BIT Mesra, Ranchi and washed for removing adhered dust particles. The washed seeds were dried in the oven at 60 °C for 48 h, and then it is crushed for making powder form. The powder was sieved and coarse particle obtained were milled again for preparing the homogeneous powder for various characterization and calcinations at 300 °C for 3 h. The schematic diagram for preparation of bio-adsorbent has been shown in Fig. 1.

2.4 Characterization of bio-adsorbent

Field emission scanning electron microscopy-energy dispersive X-ray spectroscopy was recorded on Sigma-300 with EDX (Ametek). Fourier transform infrared spectrum of the prepared bio-adsorbent using Shimadzu (Corporation, Japan; IR-Prestige 21). For phase identification, X-ray diffraction (XRD) pattern was recorded using a Rigaku, Smart Lab 9 kW diffractometer (Japan). The elemental analysis is by ICP-OES in Perkin Elmer, Optical 2100DV ICP-OES (USA).

2.5 Experimental design

Design Expert 6.0.8 has been used to design the experiments for varying different parameters. The experimental planning has been done using Box-Behnken design within RSM to





optimize the parameters in order to maximize the % removal of heavy metal present in water. Within this methodology, a 3-factor, 3 levels Box-Behnken design have been useful to study the collective outcome of three constraint pH, adsorbent doses, and concentration of heavy metal on the % removal. The diversity of the different parameters has been given in Table 1.

As per the experimental planning given by DOE, 17 experiments have been conducted and result has been fed to DOE for further analysis. The outcome of the statistical analysis is a fitted second-order polynomial in the form given below.

$$Y = C_0 + \sum C_i X_i + \sum C_{ii} X_{ii}^2 + \sum C_{ij} X_i X_j + \in,$$
(2)

where Y is the forecast yield, C_0 is constant and C_i , C_{ii} , and C_{ij} are the linear constant, quadratic constant, and the cross-product constant, respectively.

3 Result and discussion

3.1 FTIR analysis

By using FTIR spectral analysis, the important functional groups on the surface of bio-adsorbent were analyzed and shown in Fig. 2. By observing the figure, it can be observed that, at wavenumber 3279 cm^{-1} , the incorporation peak is linked with a hydroxyl group stretching vibration, and on other hand, wavenumbers around 2927 cm⁻¹ and 2364 cm⁻¹ observed for C-H aliphatic stretching vibration [14]. It is also observed that the carboxyl double bond the peak seen at 1643 cm⁻¹. Kim et al. 2015 has observed that the carboxyl group plays a vital role for cations metal absorption. The wave-number 1007 cm⁻¹ was observed for the hydroxyl group and the peak at 929 cm⁻¹ shows the carbon–carbon associates in the bio-carbon. The existence of polar groups on the bioadsorbent plane is possible to supply the significant cation substitute ability to the absorbent [34].

3.2 XRD analysis

The X-ray diffraction patterns of different waste seeds biochar powder were shown in Fig. 3. As seen in the figure, the crystalline phase availability was very small in the M, JF, and JP calcined at 400 $^{\circ}$ C for 3 h, and the large amount of amorphous

Table 1	Optimization	variables with	independent test
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Variables	Unit	Range and level			
		Low	Medium	High	
pН	-	5	7	9	
Adsorbent doses	ppm	40	60	80	
Concentration	ррт	1	1.5	2	



Fig. 2 FTIR spectra of bio-sorbent material of *Mangifera indica* (MF), *Artocarpus heterophyllus* (JF), and *Schizizium commune* (JP) calcined at 400 $^{\circ}$ C for 3 h



Fig. 3 XRD spectra of bio-sorbent material of *Mangifera indica* (MF), *Artocarpus heterophyllus* (JF), and *Schizizium commune* (JP)

part. The crystalline parts of maximum calcite (Cc) and plagioclase (Pl) and also different compositions of carbon are in biochar. The various peaks present in biochar at theta angles such as 14.89, 17.45, 22.87, and 37.97 [32]. The similar peaks found in biochars corresponded to calcite (Cc) and plagioclase (Pl) due to the alkaline oxyhydroxides formed during biochar fabrication [10], [30].

3.3 Kinetics study and equilibrium isotherm

In this research work, the arsenic removal percentage is observed, and by using three bio-adsorbents (MF, JF, and JP), it is primarily reported as shown in Fig. 4. The figure shows that the amount of adsorption of arsenic is varied with time of reaction proceeded. The arsenic adsorption gets slow down after half an hour about 70%, and after that enhanced the removal of arsenic using biochar of M (93.64%), JF (92.8%), and JP (92.64%). All these three adsorbents tested, i.e., MF, JF, and JP in 90 min duration, and at room temperature keeping pH 8.4, the experiment performed for a 120-min duration. To predict the adsorption kinetics, the four models have been applied using all three catalysts [19].

As shown in Table 2, the different models of describes the experimental data satisfied with high coefficients with comparison of recent studies. In case of the entire three bio-sorbents, Freundlich model ($R^2 = 0.99$ MF, $R^2 = 0.98$ JF, $R^2 = 0.98$ JP) and pseudo-first-order reaction kinetic ($R^2 = 0.98$ MF, $R^2 = 0.97$ JF, $R^2 = 0.97$ JP) well explained the adsorption and reaction kinetics.

Regression coefficients shown in a comparative of different kinetic models are presented in Table 3. It can be found from the table that the current and earlier study showed with regression coefficients and constants such as date seed [10], starch maghemite [20], Delonix seed biochar [10] amaltash seed biochar [9], and seed [9].

To observe the premium adsorption isotherm, the bioadsorbents MF, JP, and JF were selected for adsorption and investigational information. The mechanism of adsorption isotherm was explained by using Langmuir, Freundlich, and Tempkin model isotherms which has mentioned as above in equation (See supplementary materials). The value of R^2 is the indicative of the most excellent possible isotherm [21]. Table 3 shows the outcome of isotherm studies the entire three models, and graphically it was presented in Fig. 4 (B, C, D). From Table 3, it can be seen that the value of R^2 in Freundlich isotherm model is 0.98 for MF, 0.98 for JF, and 0.98 for JP seed catalyst. Other studies also show the similar results of the various coefficients [22].

From the Table 3 summary of equilibrium constraint of various models and their regression constants, the current investigation gives better results showed than recent researchers such as maghemite [20], date seed [10], and Delonix seed biochar [10].

3.4 Effects of various parameters on arsenic removal

The obtained results in terms of % removal for all the individual experiments are analyzed in the Design of Expert software. The enhanced investigation also performed for capitalization on the percentage removal at optimal values of procedure constraint and is present in the following subsections. The progression constraints (pH, bio-adsorbent doses, and concentration of arsenic metal) have been various in three different stages for Fig. 4 Effect of time on the % arsenic removal (A) and adsorption model (B, C, D) for arsenic removal from aqueous solution by the seed bio-sorbent of *Mangifera indica* (MF), *Artocarpus heterophyllus* (JF), and *Schizizium commune* (JP)



Table 2 Summary of kinetic of arsenic adsorption on seeds biochar of *Mangifera indica* (M), *Artocarpus heterophyllus* (JF), and *Schizizium commune* (JP)

Kinetic model	Parameters	M (present study)	JF (present study)	JP (present study)	Starch magh- emite (Siddiqui et al. 2020)	Delonix seed biochar [10]	Seed [31]
Pseudo-first	$k_1 (\min^{-1})$	0.083	0.076	0.074	0.06	0.101	0.038
order	$q_e (\mathrm{mg/g})$	5.87	6.52	7.33	2.76	4.79	0.0033
	R^2	0.985	0.979	0.972	0.60	0.921	0.9228
Pseudo-second	k_2 (g/mg. min)	0.0082	0.0036	0.00045	9.16	0.212	0.087
order	$q_e (\mathrm{mg/g})$	2.81	3.51	7.75	1.03	1.675	0.753
	R^2	0.741	0.442	0.042	0.99	0.96	0.9837
Intra-particle model	$\begin{array}{c} K_{\rm ipd} \\ (\rm mgg^{-1}\ min^{-1/2}) \end{array}$	0.229	0.21	0.217	0.03	0.274	2.01536
	$C (mg g^{-1})$	0.148	0.115	0.216	0.62	0.698	0.17576
	R^2	0.95	0.926	0.927	0.90	0.78	0.6254
Elovich model	β (g mg ⁻¹)	47.62	1.314	1.13	9.93	0.3775	6.313
	$\alpha(\text{mggmin}^{-1})$	1.94	0.030	0.0094	9.76	1.4257	5.44×10^{-6}
	R^2	0.886	0.966	0.984	0.81	0.93	0.6202

design optimizes study, and 17 different experiment runs have been investigated. For each experiment, the percentage removal has been estimated and fed to the DOE software for additional analysis. DOE guesses the model quadratic equation which communicates the independent variables (pH, bio-adsorbent doses, and concentration

Isotherm model	Parameters	M (present study)	JF (present study)	JP (present study)	Starch maghemite (Siddiqui et al. 2020)	Date seed [10]	Delonix seed biochar (Pal al. 2021)
Langmuir model	b_o (L/mg)	0.165	0.094	0.195	17.5	2.93	2.93
	$q_m (\mathrm{mg/g})$	15.29	54.6	21.74	7.15	3.2	6.2
	R^2	0.248	0.041	0.205	0.98	0.95	0.92
Freundlich model	$K_f(mg/g)$	0.365	29.25	0.360	13.8	22.43	11.09
	1/n	1.044	2.11	1.059	0.512	1.23	1.07
	R^2	0.99	0.992	0.984	0.93	0.90	0.907
Temkin model	b_T (J mol ⁻¹)	258.37	214.95	258.66	0.94	1.32	1.22
	K_T (L g ⁻¹)	0.0256	0.0338	0.0247	1.23	30.08	24.25
	R^2	0.966	0.992	0.94	0.94	0.91	0.953

Table 3 Summary of adsorption isotherm of Arsenic on Mangifera indica (M), Artocarpus heterophyllus (JF), and Schizizium commune (JP)

of arsenic metal) and dependent variable (% removal), as shown in Eq. (2). The variation of percentage arsenic removal with adsorbent dose, pH, and initial concentration using seed biomass catalyst of M, JF, and JP is shown in Fig. 5.

The percentage arsenic removal has been calculated after each experimental run, response has been provided to the Design of Expert software for statistical analysis, and quadratic model was obtained which relates the independent variables with dependent variable for a mango seed catalyst as shown in below model in Eq. (3). For JP catalyst, the model (d) was developed which is significant as it shows a very high value of model *F*-value (82.20). The DOE predicts the value of % removal for each run after analysis which is plotted verses actual experimental data and shown in Fig. 5 III (c), which shows a correct match with the value of R^2 is 0.99 between these data; hence, model is called significant.

The variation of removal percentage of arsenic by using different biochar was shown in Fig. 5 (I, II, III). Effects of interaction time (0-120 min) on the adsorption of arsenic by the M, JF and bio-adsorption are showed in Fig. 6.

 $\% removal = +59.79 - 9.53pH + 0.592CD + 25.52C + 0.97pH^{2}$ $-0.0019CD^{2} - 4.025C^{2} + 0.0015pH * CD - 0.15pH * C - 0.171 * CD * C$

In Fig. 5 I (c), the curve fitting between predicted verses actual is shown. Similarly, all experiments have been performed with JF catalyst and model (4) found as shown below:

The sorption percentage also increases and shows a linear behavior in between 15 and 80 min. The sorption quickly takes place and is generally measured by the amount of

%removal = $+69.07563 - 2.12625pH - 0.21887CD + 13.015C + 0.43313pH^{2} + 0.0015CD^{2} - 2.03C^{2} + 0.021pH * CD - 0.675pH * C - 0.0325 * CD * C$

JF catalyst is used to develop this model which shows the extent of dependency of dependent variable (% removal) on self-governing parameters. The model *F*-value indicates a significant model with high value of 73.74. Another indicative of significant model is the value of R^2 which is found to be 0.99. After analysis, the DOE predicts the value of % removal for each run which is plotted versus the actual experimental data, and shown in Fig. 5 II (c) which shows a clear match between these data; hence, the model is called significant.

Experiments have been performed with JP catalyst in similar manner and found the model (5) as shown below:

heavy metal diffuses on the catalyst surface from bulk. As on reaction proceeds for long time, active sites are being drenched with more arsenic ions.

The alteration of pH of the solution has effect on adsorption process, which alters the active site ionization of useful functional group [23]. Figure 7 shows the adsorption capacity of all three catalysts by alteration of pH value within the range mentioned and found that the percentage removal is strongly depending on pH value. It was observed that the arsenic removal increases by increasing the pH value to 8.4 from 5 and then it declines for higher value. Figure

%removal =
$$+60.042 - 5.587pH + 0.031CD + 21.85C + 0.564pH^2$$

 $-0.0014CD^2 - 3.775C^2 + 0.018pH * CD - 0.262pH * C - 0.0112 * CD * C$



Fig. 5 Shown the arsenic removal with adsorbent dose, pH and concentaration of aatalyst (I, (a), (c), (b)) MF-bioadsorbent, (II, (a), (c), (b)) JF-bioadsorbent and (III, (a), (c), (b)) JP-bioadsorbent





shows the maximum percentage removal was observed at pH value of 8.4. Figure 5 showed the removal efficiency continuously enhancing as raising the pH up to 8.4. Therefore, all the experiments were further performed at pH 8.4.

This adsorption behavior of arsenic at various pHs also be the survival of arsenic in various oxyanions appearance. A similar study was done by Issa and coauthors concluded that





Fig. 7 Influence of solution pH on arsenic % removal (contact time=90 min, pH=8.4, initial concentration=2.5 mg.L⁻¹, agitation speed=400 rpm, room temperature)

Fig. 8 Influence of seeds bio-sorbent dosage on arsenic % removal (contact time=90 min, pH=8.4, initial concentration=2.5 mg.L⁻¹, agitation speed=400 rpm, room temperature)



Table 4	Comparative study
of arsen	ic removal with other
paramet	ers

Adsorption capacity (mg/g)	рН	Adsorption time (hrs)	% Removal	Reference
6.2	3–8	2	95	Pal et al. [10]
3.2	3–8	2	93.8	Pal et al. [10]
2.6	2-10	5	79	Mohan et al. [15]
1.28	2–9	8	87	Aregawi et al. [31]
1.42	3–9	1.5	91	Giri et al. [9]
1.926	3-12	2	94	Current study
1.487	3-12	1.5	93	Current study
1.934	3–12	1.5	92	Current study
	Adsorption capacity (mg/g) 6.2 3.2 2.6 1.28 1.42 1.926 1.487 1.934	Adsorption capacity (mg/g)pH6.23-83.23-82.62-101.282-91.423-91.9263-121.4873-121.9343-12	Adsorption capacity (mg/g)pHAdsorption time (hrs)6.23–823.23–822.62–1051.282–981.423–91.51.9263–1221.4873–121.51.9343–121.5	Adsorption capacity (mg/g)pHAdsorption time (hrs)% Removal menval6.23–82953.23–8293.82.62–105791.282–98871.423–91.5911.9263–122941.4873–121.592

the pH of synthetic wastewater is very useful parameters for adsorption.

The effect of various doses on the adsorption of arsenic by the composite is presented in Figs. 5, 8, and 9. The highest arsenic removal ~ M (94%), JF (93%), and JP (92%) was recorded by 60 mg any other changes there was no effect on percentage adsorption. The chitosan-magnetic graphene oxide showed a high adsorption capacity (90.91 mg/g) at favorable neutral pH and superior removal efficiencies as high as 89% within 50 min [24]. Xu and co-authors investigated the maximum adsorption capacity arsenic was around 7.72 mg g⁻¹ by using iron-based economical and eco-friendly biochar [25].

Study shows better removal in less adsorption time and other comparative study on the effect of various parameters has been presented in Table 4. Numerous researchers have presented similar work with different bio-adsorbent and results are well matched with literature, date seed [10], delonix seed biochar [10], wood char [26], moringa stenopetala [31], almond shell [33], and amaltash seed biochar [9].

4 Conclusion

Synthesized bio-sorbents from the seeds of mango, jack fruit, and java plum well suited for the arsenic removal from the simulated wastewater. Seeds bio-adsorbent of *Mangifera indica*, *Artocarpus heterophyllus*, and *Schizizium commune* removed 94%, 93%, and 92% respectively within one and half hour duration of batch experiment at pH 8.4 and room temperature. The arsenic (As III) adsorption behavior of the seed's biochar was well fitted in terms of Freundlich isotherm, with the optimum adsorption capacity 0.365, 29.25, and 0.360 mg g⁻¹ in 90 min for MF, JF, and JP, correspondingly, whereas pseudo-second order and Elovich model were able to explain the kinetic data.

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Declarations

Conflict of interest The authors declare no competing interests.

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