CONTENTS

Page No.

List of Tables	vii-viii
List of Figures	ix-xiii
List of Abbreviations	xiv-xv
Preface	xvi-xxi

1. INTRODUCTION	1-50
1.1 Introduction to Carpet Industry	1
1.2 Synthetic Azo Dye	4
1.3 Carpet Manufacturing At a Glance	8
□Scouring	8
Desizing	8
□Bleaching	9
Dyeing	9
1.4 Environmental and Health Hazard Due to Dye bearing wastewate	r 12
1.5 Characteristics of the carpet industries wastewater	13
1.5.1 pH Value	14
1.5.2 Temperature	14
1.5.3 Acidity	15
1.5.4 Biological Oxygen Demand	15
1.5.5 Chemical Oxygen Demand (COD)	16
1.5.6 Color	16
1.5.7 Total Suspended Solids and Total Dissolved Solids	17

1.5.8 Formaldehyde17
1.5.9 Heavy Metals17
1.5.10 Pesticides
1.5.11Pentachlorophenol19
1.5.12 Prohibited Dy Stuffs19
1.6 Status of Ground Water in Cluster 19
1.7 Literature Review22
A. Methods for Dye Degradation22
1.7.1. Non-BiologicalMethods22
1.7.2 Biological Method23
1.7.2.1 Cyanobacteria and alga24
1.7.2.2Fungi24
Laccase
Ligninolytic enzymes27
1.7.2.3 Bacteria
□Bacterial amendment
Single bacterialculture
☐ Mixed bacterial culture29
□Mutant bacterial strains
1.7.3 Enzyme Azoreductase
1.7.4 Genetically Modified Bacteria44
1.7.4.1Efficiency of Genetically Modified Bacteria45
1.7.5 Mechanism of Azo Dye Degradation46
1.7.5.1 Reduction of Azo bond48
1.7.5.2 Mechanism of carcinogenicity49
1.7.5.3 Mineralization of Intermediate
1.8 OBJECTIVE OF THE PROPOSED WORK

2.EXPERIMENTAL	L
2.1 Collection and Characterization of Dye Wastewater51	
2.2 Collection of soil sample from dye contaminated sites52	2
2.3 Isolation, Screening and Identification of Dye Degrading Bacte53	;
2.3.1 Media and Chemical52	3
2.2.2 Isolation of bacterial strains54	4
2.2.3 PCR amplification and DNA sequencing of the 16S rRNA gene6	60
2.2.4 Nucleotide Sequence Accession Number	1
2.3 Inoculum Preparation62	
2.4 Optimization of various parameters for efficient degradation of dye (Batch Mode under Static condition)	2
2.4.1 Decolorization at static and shaking conditions	3
2.4.2 Effect of pH6	3
2.4.3 Effect of Temperature	4
2.4.4 Effect of initial dye concentration6	4
2.4.5 Effect of various carbon and nitrogen sources6	4
2.4.6 Investigation on capability of isolated microbe for repeated use6	5
2.5 Preparation of cell-free extract and enzyme assay6	5
2.6 Phytotoxicity studies	6
2.7 Analytical Investigation6	7
2.7.1 UV-VIS Spectrophotometer	7
2.7.2 HPLC and FTIR analysis	7
2.7.3 GC-MS analysis	8
2.8 Rectors Studies	9
2.8.1 Procedure for isolation of bacterial Consortia BC1 from soil used in aerobic dye degradation	9
2.8.2 Optimization of various parameters for efficient degradation	0
2.8.3 Experimental Set up7	0
2.8.3.1 Biodegradation through Attached culture Fixed bed Reactor7	'1

2.9 Analytical Technique Used	76
2.8.3.3 Packing Material	75
2.8.3.2 TiO ₂ coated clinkers for adsorption	74

3. Biodegradation of Navy N5rl-1 Carpet Dye By *Staphylococcus Saprophyticus* Strain Bhuss X3

3.1 INTRODUCTION	77
3.2 Result and Discussion	
3.2.1 Bacterial identification by Morphological, Biochemical an	d 16S rRNA
gene sequencing	/8
3.2.2 Parameter Optimization for Degradation Studies 80	
3.2.2.1 Effect of Agitation	80
3.2.2.2 Effect of pH	82
3.2.2.3 Effect of Temperature	83
3.2.2.4 Effect of initial dye concentration	84
3.2.3 Analytical investigation85	
3.2.3.1 UV–Vis spectral analysis	85
3.2.3.2 HPLC analysis	
3.2.3.3 FTIR Analysis	87
3.2.4 Phytotoxicity study	
3.3 CONCLUSION	90

4. Biodegradation Of C.I. Acid Red 1 By Indigenous Bacteria Stenotrophomonas Sp. Bhussp. X2 Isolated From Dye Contaminated SoiL

4.1 INTRODUCTION	
4.2 Result and Discussion	92
4.2.1 Isolation, screening and identification of bacterial strain	
4.2.2 Optimization of important operating parameter	94
4.2.3 Repeated applicability test of isolated bacteria	101
4.2.4 Enzyme Analysis	102
4.2.5 Analytical investigations	103
4.2.6 Degradation pathway	107
4.2.7 Phytotoxicty study	110
4.3 CONCLUSION	

5. Isolation Of Aerobic Mixed Consortium For Enhanced Biodegradation.

5.1 INTRODUCTION	112
5.2. Results and Discussion	115
5.2.1 Identification of Bacterial Consortia (BC1)	115
5.2.2 Parameters optimization	116
5.2.2.1 Effect of Shaking and Static Condition	117
5.2.2.2 Effect of pH	118
5.2.2.3 Effect of temperature	120
5.2.2.4 Effects of initial dye concentration	122
5.2.3 Kinetics of the dye decolorization of Scarlet 4BS	124
5.2.3.1 Determination OF K _{MAX} AND V _{MAX}	125
5.2.3.2 Effect of pH	129
5.2.3.3 Effect of dye concentration	130
5.2.3.4 Effect of initial glucose concentration and glucose to micro	obe rati.131
5.3 Effluent Degradation Bioassay	133
5.3.1 Possible pathway of Acid Red B	137
5.4 Phytotoxicity studies	141
5.5 CONCLUSION	143

6. Integrated Treatment of Carpet Effluent Using Packed Bed Reactor

6.1INTRODUCTION 144
6.2 Result and Discussion147
6.2.1. Cultivation, Enrichment and Analysis microbial Consortium148
6.2.2. Morphology of Biofilm grown on klinkers149
6.2.3. Morphology of TiO ₂ coated clinkers by using SEM analysis150
6.2.4 Performance of Aerobic Packed Bed Reactor for Dye Decolorization152
6.2.4. Adsorption of Biologically Treated Dye Effluent154
6.3 Analytical Investigation155
6.4 Phytotoxicity investigation156
6.5 Conclusion 157
7. COCLUSION AND FUTURE WORK158
REFERENCES 163
List Of Publications
List Of Paper Presented in Conferences

Carriculum Vite

	Description	Page No.
1.1	Biodegradation of dye by single strain and mixed culture of bacteria	32-38
1.2	Characteristics of enzyme azoreductase isolated from various microorganism	41-43
2.1	Showing physico-chemical characteristics of dye wastewater collected from different sites from Carpet Industries, Bhadohi and Uttar Pradesh, India	52
2.2	Characteristics of Packed Bed Reactors	72
3.1	Characteristics of isolated bacterial starin <i>Staphyolococcus saprophytic</i> BHUSS X3	79
3.2	Phytoxicity Study of Navy N5RL1 dye and its degradation metabolites	90
4.1	Effect of various carbon and nitrogen sources supplemented with MSM media ondecolorization by isolated bacteria <i>Stenotrophomonas</i> sp. BHUSS X2	100
4.2	Effect of various enzyme activities before and after decolorization of RED G dye by isolated bacteria <i>Stenotrophomonas sp.</i> BHUSSp X2	102
4.3	GC-MS spectra of degraded metabolites of Acid Red G	109
4.4	Phytotxicity Study of Acid Red G and its metabolites formed after biodegradation	110
5.1	GC-MS spectra of intermediates metabolites of degradation of	140-

Scarlet 4BS using BC1. 141

5.2	Phytotoxicity	Study	of	Scarlet	4BS	dye	and	its	degradation	142
	metabolites (S	orghum	vu	lgare)						

- 6.1 Acession numbers of 16S rRNA gene sequence of four isolated 149 bacterial strains submitted to NCBI database
- **6.2** Phytotoxicity of textile industrial wastewater and its degraded 157 products for *and Phaseolus mungo*

Figure No.	Description	Page No.
1.1	Showing the various steps of Carpet Manufacturing Unit	7
1.2	Carpet Manufacturing Unit at a Glance	11
2.1	Geographical location of the study site.	53
2.2	Showing the step for isolation of bacterial strains from dye contaminated soil.	56- 57
2.3	Showing the different bacterial strains isolated from different dye contaminated soil.	58-60
2.4	Schematics Diagram of Attached culture Fixed bed Reactor (Bioflim Packed Bed Reactor).	73
2.5	Schematics Diagram Fixed Bed TiO ₂ Coated Reactor.	74
2.6	Photograph of Reactor (a) Attached culture fixed bed Reactor (b) TiO_2 coated clinkers packing for chemical oxidation.	75
3.1	The evolutionary history of bacterial strain <i>BHUSS X3</i> was analysed by using the Neighbour-Joining Method, Bootstrap values (n=1000) are indicated at the nodes and scale bars represent 0.01 substitution/site; the sequences have been retrieved from NCBI database, showing the phylogenetic relationships of <i>Staphylococcus saprophytic BHUSS X3</i> with <i>different strain of</i> <i>Staphylococcus</i> and other species of genus <i>Pseudomonas and</i> <i>Bacillus</i> .	80
3.2	(a) Effect of Static/Shaking condition for efficient decolorization of dye Navy N5RL1	81
3.2	(b) Effect of pH on dye Navy N5RL1 decolorization by <i>Staphylococcus saprophytic</i> BHUSS X3 optimized static culture	82

condition at temperature 35 $^{\circ}\mathrm{C}$ and 100 mg dye concentration.

3.2	(c) Effect of temperature for decolorization of dye Navy N5RL1 by <i>Staphylococcus saprophytic</i> BHUSS X3.	83
3.2	(d) Effect of initial dye Navy N5RL1 concentrations (100-1000 mgl ⁻¹) under optimized static culture condition at pH 8.0, 35 °C.	84
3.3	UV-Vis spectrophotometer of Navy N5RLI (100 mg/l) biodegraded by <i>Staphylococcus saprophytic</i> BHUSSX3 before and after optimized condition at T = 35 °C, pH = 8.0, bacterium = 2.6×106 cells/ml.	85
3.4	(a) HPLC analysis for dye degradation of Dye control Sample	86
3.4	(b) HPLC analysis for dye degradation of degraded sample	87
3.5	(a) FTIR analysis of Navy N5RL Before degradation.	88
3.5	(b) FTIR analysis of Navy N5RL After degradation.	89
4.1	Phylogenetic tree: Neighbor joining phylogenetic tree based on 16S rRNA gene sequence showing the relationship between isolated bacterial strain <i>Stenotrophomonas</i> sp. BHUSSp X2 and other relatives within genus.	93
4.2	Effect of shaking and static condition on decolorization of C.I. Acid Red 1 dye by isolated bacterial strain in minimal media.	96
4.3	Effect of pH on decolorization of C.I. Acid Red 1 by strain BHUSSp X2 in minimal media. The reaction was performed in static condition with a concentration of dye 200 mg/l at temperature 35 °C. Error bars present p< 0.05 .	97
4.4	Effect of various temperature on the decolorization of C.I. Acid Red 1 by isolated bacterial strain <i>Stenotrophomonas</i> sp. BHUSSp X2 in minimal media. Conditions: pH 8; under static condition. Error bars present p< 0.05.	98
4.5	Effect of initial dye concentration on the decolorization of C.I. Acid Red 1 by isolated bacterial strain <i>Stenotrophomonas</i> sp.	99

BHUSSp X2 in minimal media. Conditions: pH 8; temperature 35°C; under static condition. Error bars present p< 0.05.

- **4.6** Repeated Use of isolated bacterial strain BHUSS X2. Error bars **101** present p< 0.05.
- UV-visible spectra of C.I. Acid Red 1 and after decolorization by 104 isolated bacterial strain BHUSSp X2 at optimized condition pH 8, Temperature 35 °C, Static condition and 100 mg/l dye concentration.
- **4.8** HPLC analysis of (a) Dye control sample (b) metabolites **105** produced after 6h.
- **4.9** FTIR spectrum of (a) Dye control sample (b) metabolites produced after 6h. **106**
- **4.10** Pathway of degradation of Acid Red 1 dye by isolated bacterial **108** strain BHUSSp. X2 by GC-MS.
- 5.1 Phylogenetic tree of Bacterial Consortia (BC1) based on 16S 116 rRNA gene sequences wherein the bar indicates the Jukes-Cantor evolutionary distance. Numbers at nodes indicate percentage bootstrap value.
- (a) Effect of different Shaking and Static condition on efficiency 117 of degradation of ACID RED G dye by mixed Consortia BC1 (dye concentration 50 mg/l and bacterium=1.5×10⁶ cells/ml)
- (b) Effect of different Shaking and Static condition on efficiency 118 of degradation of Scarlet 4BS dye by mixed Consortia BC1 (dye concentration 50 mg/l and bacterium=1.5×10⁶ cells/ml).
- (a) Effect of different pH condition on efficiency of degradation of 119 RED G dye by mixed Consortia BC1 (dye concentration 50 mg/l, bacterium=1.5×10⁶ cells/ml and temperature 35 °C under Shaking Condition 120rpm)
- (b) Effect of different pH condition on efficiency of degradation of 120
 Scarlet 4BS by mixed Consortia BC1 (dye concentration 50 mg/l and bacterium=1.5×10⁶ cells/ml, temperature 35 °C under Shaking Condition 120rpm)

- (a) Effect of different temperature condition on efficiency of dye
 degradation of Red G by Mixed Consortia BC1 (dye concentration
 50 mg/l and bacterium=2.6×10⁶ cells/ml under Shaking Condition
 120rpm)
- (b) Effect of different temperature condition on efficiency of dye
 degradation of Scarlet 4BS by Mixed Consortia BC1 (dye concentration 50 mg/l and bacterium=2.6×10⁶ cells/ml under Shaking Condition 120rpm).
- (a) Effect of different initial dye concentration on rate of 123 degradation of Red G by Mixed Consortia BC1 (bacterium=2.6×10⁶ cells/ml and temperature 35 °C under Shaking Condition 120rpm).
- (b) Effect of different initial dye concentration on rate of 124 degradation of Scarlet 4BS by isolated Mixed Consortia BC1 (Bacterium=2.6×106cells/ml and temperature 35 °C under Shaking Condition 120rpm).
- 5.6 (a) Graph of $\ln(S/S_0)$ versus time for the degradation of Scarlet 127 4BS dye using BC1.
- 5.6 (b) Estimation of activation energy of scarlet dye from graph 128 $ln(K_0M)$ versus 1000/T
- 5.6 (c) A double reciprocal plot of initial rate of dye degradation (1/V) 129 and scarlet dye concentration (1/S).
- **5.6** (d) Effect of pH on rate constant at 35 °C, 50 mg/l dye **130**
- (e) Effect of dye concentration on rate constant at 35 °C and 50 131 mg/l dye
- (f) Effect of glucose concentration on rate constant at 35 °C and 50 132 mg/l dye.
- (g) Effect of substrate to microbe weight on rate constant 35 °C 133 and 50 mg/l dye

- 5.7 (a): UV-Vis spectrophotometer of RED G (50 mg/l) biodegraded 134 by BC1 before and after optimized condition at T=35°C, pH=8.0 bacterium= 1.5×10⁶ cells/ml at Shaking Condition (120 rpm).
- 5.7 (b): UV-visible spectra of Scarlet 4BS by mixed consortia before 135 and after degradation under optimized condition i.e. temperature = 35 °C, pH= 8.0, bacterium=2.6×10⁶ cells/ml and shaking condition (120 rpm).
- **5.8** HPLC analysis for dye degradation (Acid Red G) a) Dye control **136** Sample b) Degraded sample.
- 5.9 HPLC analysis for dye degradation (Scarlet 4BS) a) Dye control 136 Sample b) Degraded sample.
- 5.10 Proposed pathway for the degradation of Scarlet 4BS by isolated 138-bacterial consortia (BC1) on the basis of identified metabolites, 159 enzyme activity and utilization profile of different substrates.
- 6.1 Scanning Electron Micrographs of support material clinkers 150 immobilized bacterial strain (a) Control Clinkers (without bioflim)
 (b) Bioflim formation on clinkers.
- 6.2 Scanning Electron Micrographs of Support material clinkers 151 coated $TiO_2(a, b)$ before degradation (c) After Degradation.
- 6.3 Performance of Packed Bed Bioflim Bioreactor operations with 153 the function of different concentration of dye.
- 6.4 Performance of biofilm Packed Bed Bioreactor operations with the 153 function of COD removal.
- 6.5 Combined treatment of dye wastewater using (Packed Bed Bioflim 155 Bioractor + TiO₂ Coated Chemical Oxidation Reactor
- 6.6 Variations in the UV-Visible spectra of inlet and outlet samples 156 with the function of time in Packed Bed Bioflim Bioractor + TiO_2 Coated Chemical Oxidation Reactor.

LIST of ABBREVIATIONS

COD	:	Chemical Oxygen Demand
TSS	:	Total Suspended Solids
TVSS	:	Total volatile suspended solids
MPN	:	Most Probable Number
HPLC	:	High Performance Liquid Chromatography
FTIR	:	Fourier transform infrared spectroscopy
DCIP	:	Dichloro-phenol indophenol
NADH	:	Nicotinamide adenine dinucleotide
NADPH	:	Nicotinamide adenine dinucleotide phosphate
AQDS	:	Anthraquinone-2, 6-disulfonate
MSM	:	Mineral Salt Media
NCBI	:	National Center for Biotechnology Information
BLAST	:	Basic Local Alignment Search Tool
CDRI	:	Central Drug Research Institute
BMM	:	Basal Mineral Medium
NADH- DCIP		Nicotinamide adenine dinucleotide Dichloroindophenol
MSM	:	Mineral Salt Media
DCIP	:	Dichloro-phenol indophenols
GC-MS	:	Gas chromatography-mass spectrometry
PCR	:	Polymerase chain reaction
DNA	:	Deoxyribonucleic acid
rRNA	:	Ribosomal ribonucleic acid
IAA	:	Indole Acetic Acid

NaCl	:	Sodium chloride
NaOH	:	Sodium hydroxide
HCl	:	hydrochloric acid
ABTS	:	2,2'-azino-bis(3-ethylbenzothiazoline-6- sulphonic acid)

PREFACE

Carpets were probably first made by nomadic peoples to cover the earthen floor in their tents. Carpets were known in India as early as 500 B.C. references to woven mats and floor coverings can be found in ancient and medieval Indian literature. Indian Carpet Industry comprises of Hand knotted carpet, Hand tufted and Machine made carpet. However, Indian hand-knotted carpets receive worldwide acclaim. Perhaps the most famous carpet-producing region in India is the "carpet belt" of Uttar Pradesh. This belt is located near Varanasi in the south eastern corner of U.P., one of the state's poorest regions, and is capable of producing virtually every variety of carpet. Production and trade activities are primarily concentrated in the Mirzapur-Bhadohi area and extend outwards to surrounding districts. This industry is endowed with a fully large and diversified production base with an estimated 3 lakhs looms. Providing employment to 1.6 million as carpet weavers and others engaged in allied activities. Bhadohi-Mirzapur belt in U. P. accounts for around 80% of activities pertain to production & export. The Industry is export oriented and positive Net Foreign Exchange (NFE) earner contributing to the growth of Indian Economy in general and textile industry in particular Bhadohi.

Changing fashion trends and through away attitude as resulted into drastic increase in the use of azo dyes due to their bright shades and lasting fastness property. More than ten thousand sets of azo dyes are currently being used amounting to total production of approximately one million tons per annum. The industries which are major consumers of synthetic azo dyes are textile, carpet, lather, printing, food, cosmetics,

pharmaceuticals, paints etc. Due to the presence of conjugated bond and its aromatic structures, azo dyes are very stable and high resistant to light, washing and microbial attack. Synthetic dyes are classified according to chemical structure, molecular structure and their application. The effluents being generated from a dying plant is generally large in amount and complex in nature as it is the sum of dying and washing processes. Out of the coloring compound being used for dying, azo dyes are most prominent. Thus effluent coming out of these industries contain large amount of hazardous chemical and dyes and it has been as as one of the worst anthropogenic polluters. Various physico-chemical treatment techniques were attempted for treatment but they have limitation of large energy consumption and generation of secondary products (Singh and Arora, 2011, Toor et al., 2012, Liao et al., 2012 Pandey et al., 2007, Forgacs et al. 2004, Bafana et al., 2008). Although earlier investigations show that these effluents exhibits very slow degradation and are resistant to conventional biological treatment due to their complex aromatic nature. Bioremediation, utilizing microorganisms/enzymes (Ryan et al. 2005), is a potential technique for treatment of industrial wastewater. present investigation aims at complete treatment of carpet effluent using bioremediation and its integration with tertiary treatment using adsorption. Batch investigation using pure and mixed consortia indigenously isolated from contaminated soil from study area were used for the said treatment. Operating parameters such as pH, temperature, initial dye con etc were optimized during the batch studies. The most efficient mixed consortia were again tested for its efficacy in a sequential batch reactor. For the final treatment to obtain nearly 100%

degradation biologically treated effluent were sent for adsorptive treatment onto TiO2 coated clinkers, in a cylindrical reactor. The detail discussion as under:

Chapter-1 starts with the general introduction describing the carpet industries and steps involving the manufacturing of the Carpet Industries near Bhadohi, Uttar Pradesh, India. The use of synthetic dye and its effects on the environments have been discussed briefly. The physicochemical characteristics of dye wastewater produced by these industries were discussed. The wastewater coming from dye based industries contaminates nearby water bodies like river, ground water and other water ways. A review literature pertaining different methods of treatment of dye wastewater and most significant method utilized for treatment of dye wastewater were bioremediation has been presented under 'Literature Review'. The scope and objective of the present investigation have been highlighted at the end of this chapter.

Chapter-2 describes the collection of dye wastewater and dye contaminated soil sample at different sites of study area. The isolation of bacterial strain from dye contaminated soil for degradation of various dyes. This chapters also heighlights the characterization and identification of isolated bacterial strains by using morphological, biochemical and 16S rRNA gene sequence analysis. This chapter deals with the various optimizations of parameters such as pH, temperature, dye concentration, static and agitation speed condition for efficient degradation of dye. The protocol describes for the identification enzyme involved in dye degradation. The dye control sample and after degradation were characterize by using analytical techquni such as UV-Visible Spectrophotometer (UV-Spectrophotometer), Fourier Transform Infrared Spectroscopy (FTIR) and High Performance Liquid Chromatography (HPLC). The dye and dye degradation pathway were studies by using Gas chromatography–mass spectrometry (GC-MS). Phytotoxicity of dye and dye degrded metabolites were examined by using different types seeds.

Chapter-3 Biodegradation of Navy N5RL1, a widely used acidic azo dye in carpet industry was studied by bacterial strain isolated from the dye contaminated soil collected from a carpet industry premises located in Bhadohi, Sant Ravidas Nagar and Uttar Pradesh, India. The isolated strain was identified as *Staphylococcus saprophyticus* BHUSS X3 on the basis of morphological, biochemical and 16S rRNA gene sequencing analysis. The strain BHUSS X3 decolorized 95.7% of dye (100 mg/l) within 6h at optimum pH-8, temperature 35 °C, inoculum 4.0% under static condition during 24h incubation. The isolated BHUSS X 3 can treat higher concentration upto dye 1000 mg/l. The dye degradation metabolites were confirmed by analysis of degraded products using UV-Vis spectrophotometric, HPLC and FTIR technique. The phytotoxicity analysis was also conducted on *Phaseolus aureus* and enhanced seed germination was recorded.

Chapter-4 A significant proportion of xenobiotic recalcitrant azo dyes are being released in environment during carpet dyeing. The bacterial strain *Stenotrophomonas* sp. BHUSSp X2 was isolated from dye contaminated soil of carpet industry, Bhadohi, India. The isolated bacterial strain was identified morphologically, biochemically and on the basis of 16S rRNA gene sequence. The isolate decolorized 97% of C.I. Acid Red 1 (Acid RED G) at the concentration of 200 mg/l within 6 h under optimum static conditions (temperature-35°C, pH-8 and initial cell concentration-7 × 10⁷ cell/ml). Drastic reduction in dye degradation rate was observed beyond initial dye concentration from 500 mg/l

(90%) and it reaches to 25% at 1000 mg/l under same set of conditions. The analysis related to decolorization and degradation were done using UV-Vis spectrophotometer, HPLC, and FTIR, whereas, the GC-MS technique was utilized for the identification of degradation products. Phytotoxicity analysis revealed that degradation products are less toxic as compared to the original dye.

Chapter-5 in the present chapter mixed consortia bacterial starin was isolated from soil from the drain caring dying effluent of various industries. The mixed consortia were identified by 16S rRNA gene sequence analysis. The microbial consortium was dye degradation. In addition they have good bioflim forming ability and have high resistance to higher concentration developed in the laboratory and used batch methods. The mixed consortia three ifferent dye such. The degradation and dye metabolites were analysis by using UV-Spectrophotometer and HPLC.

Chapter-6 The mixed consortia of microorganism isolated from soil sample of carpet effluent fared alot better as compared to individual microorganism. Bioremediation experiment in indigenously designed and fabricated bioreactor using clinkers as packing material resultant into resign ably good decolorization upto 72%. To achieve surface discharge conditions the treated sample was sent for adsorptive treatment using packed bed reactor having TiO₂ coated clinkers as adsorbent. The decolorization upto 96% could be achieved with this integrated scheme of treatment. Phytotoxicity studies through germination of *P. mungo* seeds in this treated water was comparable to this control whereas only 66% and 26% germination was observed in case biologically treated and pure effluent, respectively.

Chapter-7 This chapter describes the salient features, main observations, key finding followed by feature works.

A consolidated list of books and Journal patents consulted during this study has been given at the end of thesis under references heading.